



ELEVATIONAL GRADIENTS DO NOT AFFECT THERMAL TOLERANCE  
AT LOCAL SCALE IN POPULATIONS OF LIVEBEARING FISHES  
OF THE GENUS *LIMIA* (CYPRINODONTIFORMES: POECILIINAE)

**Gradientes de elevación no afectan la tolerancia térmica a escala local en poblaciones  
de peces vivíparos del género *Limia* (Cyprinodontiformes: Poeciliinae)**

Rodet Rodriguez-Silva<sup>1a\*</sup> and Ingo Schlupp<sup>1b</sup>

<sup>1</sup>Department of Biology, University of Oklahoma, 730 Van Vleet Oval, Norman, OK 73019; <sup>1a</sup> [orcid.org/0000-0002-7463-8272](https://orcid.org/0000-0002-7463-8272); <sup>1b</sup> [orcid.org/0000-0002-2460-5667](https://orcid.org/0000-0002-2460-5667), [schlupp@ou.edu](mailto:schlupp@ou.edu). \*Corresponding author: [rodet.rodriguez-silva-1@ou.edu](mailto:rodet.rodriguez-silva-1@ou.edu).

ABSTRACT

One of the main assumptions of Janzen's mountain passes hypothesis is that due the low overlap in temperature regimes between low and high elevations in the tropics, organisms living in high-altitude evolve narrow tolerance for colder temperatures while low-altitude species develop narrow tolerance for warmer temperatures. Some studies have questioned the generality of the assumptions and predictions of this hypothesis suggesting that other factors different to temperature gradients between low and high elevations may explain altitudinal distribution of species in the tropics. In this study we test some predictions of the Janzen's hypothesis at local scales through the analysis of the individual thermal niche breadth in populations of livebearing fishes of the genus *Limia* and its relationship with their altitudinal distribution in some islands of the Greater Antilles. We assessed variation in tolerance to extreme temperatures (measured as critical thermal minimum (CTmin) and maximum (CTmax) and compared thermal breadth for populations of eight species of *Limia* occurring in three Caribbean islands and that occupy different altitudinal distribution. Our results showed that species analyzed had significant differences in thermal limits and ranges. Generally, species distributed in high and low elevations did not differ in thermal limits and showed a wider range of thermal tolerance. However, species living in mid-elevations had narrower range of temperature tolerance. We found no significant effect of phylogeny on CTmin, CTmax and thermal ranges among species. This study did not provide evidence supporting Janzen's hypothesis at a local scale since thermal tolerance and altitudinal distribution of *Limia* species were not related to temperature gradients expected in nature. Phylogeny also did not explain the patterns we observed. We suggest that biotic factors such as species interactions, diet specializations, and others should be considered when interpreting current distribution patterns of *Limia* species.

*Keywords:* Caribbean, elevation, species distribution, temperature.

RESUMEN

Una de las premisas más importantes de la teoría de Janzen acerca del efecto de las elevaciones en la dispersión de las especies es que, debido al bajo solapamiento en los regímenes de temperatura entre un gradiente de elevación en los trópicos, los organismos que habitan en altitudes elevadas desarrollan mayor tolerancia a temperaturas bajas mientras que aquellos que viven en zonas de baja elevación exhiben mayor tolerancia por temperaturas altas. Sin embargo, algunos estudios han cuestionado la generalidad de las premisas y predicciones de esta hipótesis sugiriendo que otros factores no relacionados a los gradientes de temperatura pudieran explicar la distribución altitudinal de muchas especies en los trópicos. En el presente estudio se someten a prueba algunas de las predicciones de la teoría de Janzen a escala local a través del análisis

de la amplitud del nicho térmico en poblaciones de peces del género *Limia* y su relación con la distribución altitudinal de estas especies en algunas islas de las Antillas Mayores. Evaluamos las variaciones en tolerancia térmica a temperaturas extremas [medidas como temperatura crítica mínima (CTmin) y máxima (CTmax)]. Además comparamos el intervalo térmico en poblaciones de ocho especies de este género que habitan en tres islas del Caribe las cuales se distribuyen en diferentes altitudes. Nuestros resultados muestran que existen diferencias tanto en los límites como en los intervalos de temperatura entre las especies analizadas. Generalmente, las especies distribuidas en altas y bajas elevaciones no muestran diferencias en sus límites térmicos y por lo general estas especies exhiben un amplio intervalo de tolerancia térmica. Sin embargo, las especies que habitan en elevaciones medias muestran un intervalo más estrecho de tolerancia térmica. Los análisis filogenéticos no explican los patrones observados en este estudio. Nuestro análisis no provee evidencia que soporte la teoría de Janzen a escala local en peces del género *Limia* ya que la tolerancia térmica y distribución altitudinal de las especies no están relacionadas con los gradientes de temperatura esperados en condiciones naturales. Sugerimos que factores abióticos tales como interacciones inter-específicas o especializaciones en la dieta, deben ser considerados para la interpretación de los patrones de distribución en peces del género *Limia*.

*Palabras clave:* Caribe, elevación, distribución de especies, temperatura.

## INTRODUCTION

Species distributions in natural systems are strongly modulated by climate, which ultimately affects both the ecology and physiology of organisms. This is particularly evident in ectothermic animals. Janzen (1967) published one of the most prominent papers in ecology that connected climatic variation across latitude and elevation, physiological adaptation and species distribution in a synthetic theory commonly referred as “Janzen’s hypothesis” (Ghalambor *et al.*, 2006; Muñoz & Bodensteiner, 2019). One of the main predictions of this hypothesis is that due to the decrease in mean annual temperature with elevation, the seasonal temperature overlap is lower in the tropics than in temperate regions. Hence, mountain passes in the tropics may represent more effective physiological barriers to dispersal than the topographical component of change in altitude (Ghalambor *et al.*, 2006). Therefore, the low overlap in temperature regimes between low and high elevations in the tropics should select for organisms with relatively narrow thermal tolerances. Janzen’s hypothesis also predicts that species develop physiological adaptations mirroring the range of ecological variation present in their surrounding area with populations living in high altitude evolving narrow tolerance for colder temperatures while low altitude populations developing narrow tolerance for warmer temperatures. Janzen’s hypothesis has been widely adopted and some studies have provided at least partial evidence at both local and global scale supporting his predictions and assumptions in both terrestrial and aquatic ectothermic organisms. For example, Pintanel *et al.* (2019) found that frog species occurring in open habitats, such as in valleys and lowland environments in general, had higher tolerance to high temperatures (CTmax) than species restricted to forest habitats, showing small climatic overlap across an elevation gradient. Moreover, Polato *et al.* (2018) provided strong evidence in support of Janzen’s hypothesis showing that tropical stream insects had noticeably narrower thermal tolerances and a lower dispersal ability than temperate species, which result in higher tropical speciation rates.

However, despite of general support of the theory, several components have never been thoroughly tested and critically evaluated across multiple taxa, potentially questioning the generality of Janzen’s theory (Ghalambor *et al.*, 2006). In addition, under Janzen’s hypothesis is unclear whether the predictions refer to individual thermal niches or species thermal niches, which in fact are determined by different factors (Hua, 2016). In fact, some studies have not found

support for Janzen's theory: in amphibians (Valdivieso & Tamsitt, 1974) and in *Anolis* lizards from Hispaniola (Muñoz & Bodensteiner, 2019) factors such as daily variation in temperature and behavioral mechanisms might cause deviations from Janzen's predictions. Furthermore, Navas *et al.* (2013) demonstrated that the effect of different microclimates within a specific biome is more relevant for species distributions than just the elevation at which certain species of amphibians may be found. McCain (2009) provided additional evidence for the effects of thermoregulation, daily temperature variability, and other climate variables such as precipitation as potential variables that could explain distribution ranges across multiple groups of vertebrates, including mammals, birds, reptiles and amphibians. In other words, Janzen's theory may have to be amended by including more complexity.

Several key features related to the geographic distribution of *Limia* make these fishes an excellent system to explore how temperature fluctuations associated to elevational gradients might be linked to dispersal. *Limia* fishes are one of the most dominant groups in freshwater ecosystems in the Caribbean with at least 19 endemic species on Hispaniola and one endemic species each occurring in Cuba, Jamaica, and Grand Cayman (Burgess & Franz, 1989; Rodríguez, 1997; Hamilton, 2001; Rodríguez *et al.*, 2020). These freshwater fishes occur in a wide distribution range occupying diverse aquatic habitats on these islands (Weaver *et al.*, 2016a).

Even though thermal regimes of Greater Antillean streams are relatively stable, geological differences among islands lead to some climate heterogeneity that can generate environmental barriers. Hispaniola, for example, has several mountain ranges of more than 2000 meters in altitude. For instance, Pico Duarte in the Dominican Republic, the highest peak in the Caribbean, reaches 3098 meters. Previous studies have shown that high elevation specialists (mainly amphibians and reptiles) have evolved on Hispaniola as a consequence of this climate heterogeneity (Wollenberg *et al.*, 2013; Muñoz *et al.*, 2014). Mountains reaching around 2000 meters can be also found in eastern Cuba (Pico Turquino) and Jamaica (Blue Mountains) with significant levels of biodiversity associated.

Although the altitudinal distribution of freshwater fish species in general is known to be considerably more constrained than in terrestrial species by several factors including for example productivity, physicochemical characteristics of the water and others (Jaramillo-Villa *et al.*, 2010; Graham *et al.*, 2014; Carvajal-Quintero *et al.*, 2015), differences in altitudinal distribution in species of the genus *Limia* can be observed in natural habitats. In the present study, we tested some predictions of the Janzen's hypothesis at the local scale through the analysis of the individual thermal niche breadth in several populations of livebearing fishes of the genus *Limia* and its relationship with their altitudinal distribution in some islands of the Greater Antilles in the Caribbean.

According to theory, we hypothesize that populations of species distributed in lowland habitats have evolved to resist higher extreme temperatures, which may be a factor limiting their dispersal into higher elevations. Conversely, populations occurring at higher elevations in mountain streams should have evolved to cope with lower temperatures, which reduce dispersal abilities into warmer habitats. Specifically, we predict that low elevation populations will be more tolerant to higher temperatures than mid and high elevation populations showing higher critical thermal maximum (CT<sub>max</sub>) and critical thermal minimum (CT<sub>min</sub>). In contrast, high elevation populations are expected to be more tolerant to lower temperatures showing lower CT<sub>max</sub> and CT<sub>min</sub> values. We also predict the thermal breadth (the range of temperatures they can tolerate) to be smaller for higher altitude fishes as result of little variability in CT<sub>max</sub>.

## OBJECTIVES

- Test some predictions of the Janzen's hypothesis at local scales through the analysis of the individual thermal niche breadth in populations of livebearing fishes of the genus *Limia* and its relationship with their altitudinal distribution in some islands of the Greater Antilles.

## MATERIALS AND METHODS

The care and use of experimental animals complied with the University of Oklahoma animal welfare laws, guidelines and policies as approved by Animal Welfare Assurance on file with the Office of Laboratory Animal Welfare under the assurance number A3240-01. Experiments were performed under the approved IACUC protocol R17-011 and specimens were collected in the field as part of surveys of the native livebearing fishes of the Greater Antilles (protocol R18-005). No fishes were euthanized nor surgical procedures were performed.

*Study area and species.* In this study we analyzed populations of some species such as *L. perugiae*, *L. vittata*, *L. yaguajali* and *L. sulphurophila* that were reported to live in low elevation, warm environments including saline coastal lagoons. We also included in the analysis other species such as *L. zonata* and *L. melanogaster* that were obtained from low to intermediate elevations and often associated to relatively cool springs. Finally, one population of *L. dominicensis* and another of *L. versicolor* collected in mountain streams at relatively high elevations were analyzed too (Table I).

Table I. List of species and populations of *Limia* included in this study with GPS coordinates and elevation in meters above mean sea level (m.a.m.s.l.) of original collecting sites. Number of individuals analyzed of each sex is also included in all cases

Species	Locality/GPS coordinates	Collection year	Sample size and sex	Elevation (m.a.m.s.l.)
<i>Limia perugiae</i>	Lake Enriquillo 18° 24' 4.61" N 71° 34' 16.61" W	2014	7 females 3 males	0
<i>Limia perugiae</i>	Las Salinas (Bani) 18° 12' 43.488" N 70° 32' 27.095" W	2011	6 females 4 males	0
<i>Limia perugiae</i>	Azufrada 18° 33' 40.212" N 71° 41' 50.928" W	2003	5 females 5 males	0
<i>Limia perugiae</i>	La Zurza 18° 23' 56.08" N 71° 34' 13.22" W	2014	5 females 5 males	0
<i>Limia vittata</i>	Cuba (Unknown coordinates)	Unknown	5 females 5 males	Unknown
<i>Limia sulphurophila</i>	Cabral (Barahona Province) 18° 14' 45.902" N 71° 13' 23.944" W	2014	6 females 4 males	26
<i>Limia yaguajali</i>	Arroyo del Agua (Jamao al Norte) 19° 37' 49.872" N 70° 26' 58.056" W	2018	5 females 5 males	46

Table I. Continuation

Species	Locality/GPS coordinates	Collection year	Sample size and sex	Elevation (m.a.m.s.l.)
<i>Limia melanogaster</i>	Roaring River 18° 17' 00.0" N 78° 03' 22.0" W	2017	9 females 3 males	71
<i>Limia zonata</i>	Rio Yuna (Bonaó) 18° 57' 33.1" N 70° 24' 32.06" W	2014	6 females 4 males	158
<i>Limia versicolor</i>	Rio Basima 18° 42' 5.605" N 70° 11' 50.387" W	2018	5 females 5 males	181
<i>Limia dominicensis</i>	Puerto Escondido 18° 19' 6.93" N 71° 34' 14.24" W	2014	5 females 5 males	399

We compared thermal breadth for the eight populations of the *Limia* species abovementioned that naturally occur in Cuba, Hispaniola, and Jamaica (Fig. 1). For a more fine-grained picture, we analyzed three additional populations of *L. perugiae*, the most widely distributed species of *Limia* on Hispaniola in order to test for local adaptation to environmental variation in temperature. All populations analyzed but *L. vittata* came from wild caught stocks that were transported into the United States and then kept in common garden conditions at the University of Oklahoma for variable periods of time (Table I). *L. vittata* specimens were obtained from aquarium stocks and have been kept in common garden conditions at a greenhouse in the Aquatic Research Facility at the University of Oklahoma for more than 10 years.

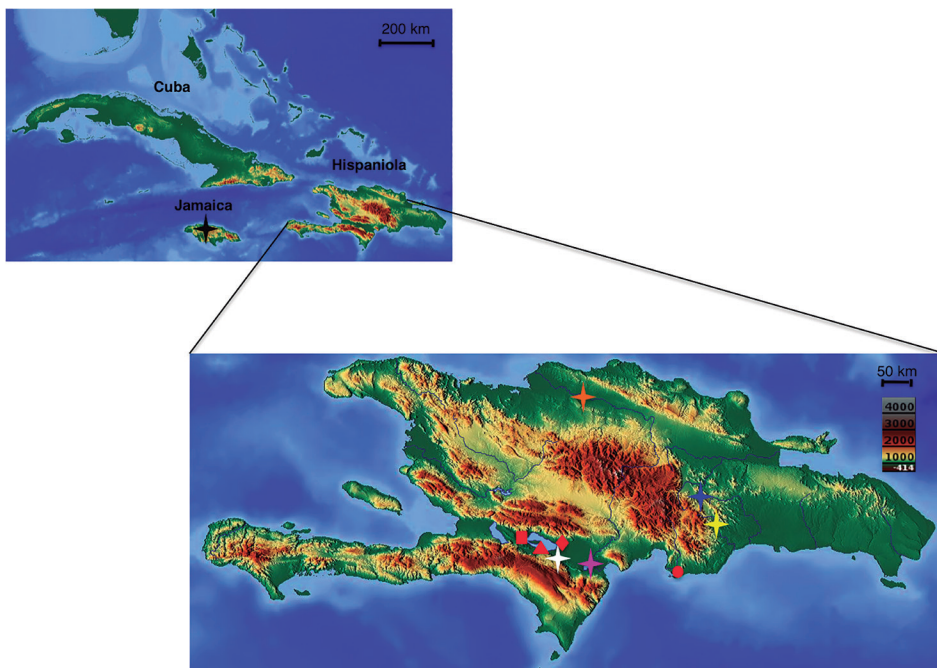


Figure 1. Geographic distribution of the populations of *Limia* (with known origins) analyzed in this study. Red rectangle: *L. perugiae* (Azufrada); Red triangle: *L. perugiae* (Lake Enriquillo); Red diamond: *L. perugiae* (La Zurza); Red circle: *L. perugiae* (Las Salinas); Orange star: *L. yaguajali*; Blue star: *L. zonata*; Black star: *L. melanogaster*; White star: *L. dominicensis*; Yellow star: *L. versicolor*; Pink star: *L. sulphurophila*.

*Laboratory methods.* We used the critical thermal method (Cowles & Bogert, 1944) to describe variation in temperature tolerance in 113 adult fish of eight *Limia* species representing a total of 11 different populations (Fig. 1). Prior to testing all fishes were acclimated to laboratory conditions with temperatures ranging between 25 °C – 27 °C for 45 days. This was the most common temperature range for the species at their origins. We tested 10 reproductively mature adult fish per population (Table I).

Fishes were individually tested under a constant increase (to determine critical thermal maximum, CTmax) or decrease (to determine critical thermal minimum, CTmin) in temperature until reaching an appropriate endpoint. The endpoint we used to determine CTmax was the pre-death thermal point at which signals of sudden onset of muscular spasms appeared (Lutterschmidt & Hutchison, 1997; Beitinger *et al.*, 2000). In the case of the CTmin the endpoint used was the absence of motion of the pectoral fins in which the fish did not start to move again even when the experimenter disturbed the fish (Fischer & Schlupp, 2009). Using this data, temperature tolerance was calculated as the arithmetic means of high and low temperatures (CTmax or CTmin) at which the endpoint was reached by individuals in the sample (Lowe & Vance, 1955). We also calculated the thermal breadth as CTmax minus CTmin for each individual. The three measures, although connected, reflect different physiological properties of the species.

Before the actual experiment fishes were not fed for 24 hours. Each fish was individually tested in a spherical 2-liter glass container. After a 10-minutes acclimation period before each trial, the container was heated using a concave heating plate at a constant rate of 1 °C/min while temperature was constantly monitored with a thermometer in order to record CTmax. Each trial was immediately stopped, and the final temperature was measured once the fish showed symptoms of sudden muscular spasms, which were characterized by disorganized and high frequency muscular movements. All fish were weighed after each trial, placed in individual tanks, and allowed to rest for at least 72 hours until application of the other extreme temperature to the same individual. Temperature exposure (CTmax or CTmin) that a fish experienced first was randomized to avoid an order effect. This was also evaluated statistically.

A similar procedure was followed to test the fish's tolerance to cold temperatures or CTmin. In this case, we placed the fish in a similar 2-liter glass container and after a 10-minute acclimation period, we continuously added cold water of 3 °C – 4 °C to the system for a rate of temperature change of 1 °C/min. CTmin was measured at the point in which fish showed total absence of movement. No mortality was associated with the trials and after the experiments all fishes were returned to their respective stock tanks.

*Climate index overlap.* Monthly maximum and minimum temperatures for the 2010–2019 period were extracted from WorldClim database for the following localities included in Table I: La Zurza, Cabral, Arroyo del Agua, Roaring River, Río Yuna, Río Básima and Puerto Escondido. We calculated the average monthly temperature for each site during the last 10 years in order to measure pairwise thermal overlap between each focal site and the other localities using Janzen's (1967) equation:

$$\text{Overlap value} = \sum_{i=1}^{12} \frac{d_i}{\sqrt{R_{ii}R_{ii}}}$$

where  $d_i$  is the thermal overlap between the focal site and each other site for the  $i^{\text{th}}$  month or the amount (in Celsius degrees) of one thermal regime that is included within the other,  $R_{ii}$  is the difference between the monthly mean maximum and minimum for the focal site and  $R_{ji}$  is the corresponding value for each other site of the study. As temperature overlap increases

the overlap value increases up to a value of 12 which is the point where thermal regimes between two sites share identical monthly maximum and minimum temperatures throughout the year.

*Phylogenetic signal.* A common caveat of studies like this is that any pattern found might not necessarily reflect adaptations but be due to species relatedness. To test for this, we used a previously-inferred phylogeny based on three mitochondrial (12S, ND2, Cytb) and two nuclear (MYH6, Rh) genes and conducted a test of phylogenetic signal to assess whether correlations in temperature tolerance among species may be due to their shared evolutionary history or to other factors (Gingras *et al.*, 2013; Kamilar & Cooper, 2013; Gilbert *et al.*, 2018; Arnaudo *et al.*, 2019). For this analysis we used Pagel's lambda ( $\lambda$ ), (Pagel, 1994) as a quantitative measure of this relationship. The Pagel's  $\lambda$  has been shown to be a very robust indicator of a correlation between ecological and evolutionary processes even for incompletely resolved phylogenies (Molina-Venegas & Rodriguez, 2017; Leiva *et al.*, 2019). We based this analysis on the phylogeny published by Weaver *et al.* (2016b), which includes most of the species used in this study.

*Data analysis.* For data analysis of thermal tolerance, we used one-way ANOVA's, after confirmation of homogeneity of variances by Levene's tests and normality by Shapiro and Wilk's tests. We used three different ANOVAs to compare the mean CTmax, CTmin and temperature ranges (breadth) among eight different populations of *Limia*. Scheffe's post hoc tests were used to make comparisons between groups to distinguish populations that differed from others in extreme thermal tolerance or temperature ranges. The lack of order effect was statistically confirmed ( $p > 0.05$ ) using t-test analyses to compare CTmax and CTmin means of fish that were tested CTmax then CTmin versus fish that were tested CTmin then CTmax.

All statistical analyses were performed in SPSS 23. We performed independent ANOVA analyses because we considered CTmax and CTmin as ecologically and evolutionary independent variables with potentially different adaptive benefits. Similar approaches that consider the effects of these variables (and also acclimation temperature) as independent have been used in other studies examining thermal tolerances in ectothermic animals (Spotila, 1972; Layne & Claussen, 1982). To further explore a potential role for local adaptation in thermal tolerance within a widely distributed species, we also compared the same variables through separate ANOVA analyses in four populations of *L. perugiae*.

In order to calculate climate index overlap we used the R package raster (Hijmans, 2020) to extract minimum and maximum monthly temperatures values to our sampled locations. Finally, analysis of phylogenetic signal was computed using the R package (R Core Team, 2013) phytools (Revell, 2012).

## RESULTS

*Analysis of phylogenetic signal.* The analysis of phylogenetic signal determined whether more similar values in CTmin, CTmax and thermal range were associated with more closely related species more often than expected by chance. None of the analyses found a significant effect of phylogeny in explaining thermal breadth among the *Limia* species studied: CTmin ( $\lambda = 6.257973e-05$ ,  $p = 1.000$ ), CTmax ( $\lambda = 0.4056982$ ,  $p = 0.78255460$ ) and thermal range ( $\lambda = 1.16242$ ,  $p = 0.362986$ ).

*Analysis of climate index overlap.* Overall, thermal overlap along altitudinal gradients among the study locations was relatively high but there was still some variation across pair of sites (Table II). Climate index overlap decreased as the differences in elevations were more conspicuous.

Particularly, Puerto Escondido (the highest collecting site included in the analysis) showed the lowest climate overlap with all other sites, which indicates that there are some differences in habitat temperatures.

Table II. Pairwise values of climate index overlap across the seven collecting sites showing different altitudinal gradients

	La Zurza	Cabral (Barahona)	Jamao al Norte	Roaring River	Río Yuna (Bonaó)	Río Básima	Puerto Escondido
La Zurza	–						
Cabral (Barahona)	11.47	–					
Jamao al Norte	10.63	11.15	–				
Roaring River	10.81	11.22	10.28	–			
Río Yuna (Bonaó)	10.29	11.39	11.59	9.96	–		
Río Básima	10.05	10.55	11.74	9.65	11.84	–	
Puerto Escondido	8.93	9.38	10.17	8.45	10.57	10.79	–

Climate index overlap can take values up to a maximum of 12 where the higher values indicate more temperature overlap.

*Inter-specific analysis of thermal tolerance.* The overall range of temperature tolerance for the eight species included in the analysis was 12°C (CTmin) to 41.2 °C (CTmax), which may be considered as a broad range for tropical fishes when considering the overall climatic stability present in the tropics in terms of temperature fluctuations. A one-way ANOVA was conducted to compare the effect of varying distribution according to elevation on the temperature tolerance under CTmin and CTmax conditions. There were significant differences in thermal limits for both CTmin (One-way ANOVA,  $F [7, 72] = 41.977, p < 0.001$ ) and CTmax (One-way ANOVA,  $F [7, 72] = 14.878, p < 0.001$ ) among species after testing 80 individuals. The highest temperature tolerance was recorded for *L. sulphurophila* with an average CTmax of 40.9 °C. This species also showed the lowest tolerance to low temperatures with an average CTmin of 16.7 °C, which might suggest this species could be adapted to live in warmer habitats. Post hoc analysis also showed that *L. sulphurophila* differed significantly from all other species in both CTmin and CTmax (Scheffe,  $p < 0.05$ ) except for *L. zonata* in CTmin (Scheffe,  $p = 0.137$ ), (Fig. 2).



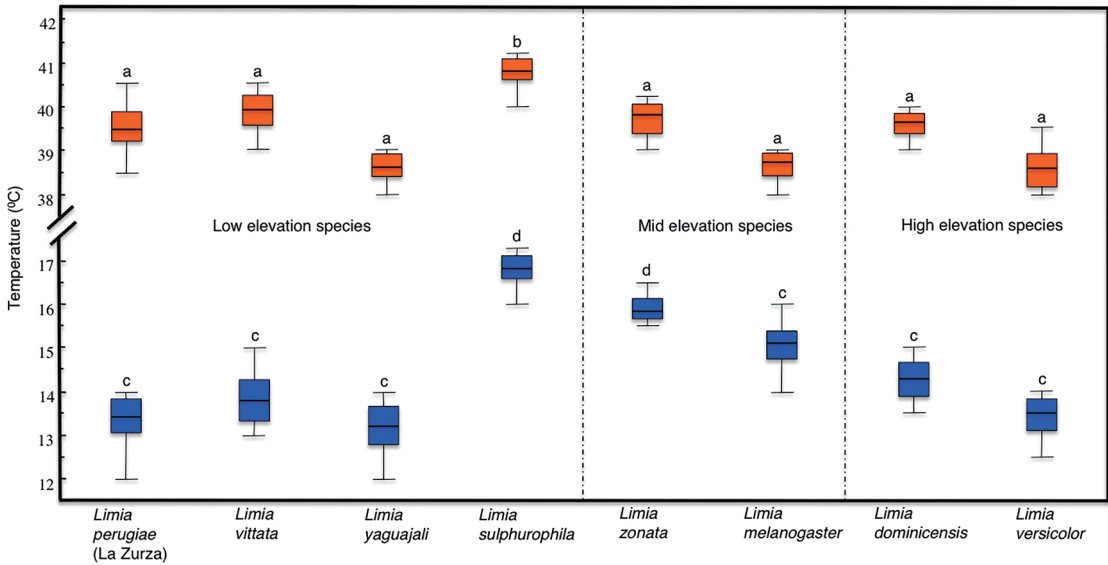


Figure 2. Thermal tolerances of the eight species of *Limia* included in this study. Species are grouped according to their elevation distribution in three groups: low elevation species (left), mid elevation species (center) and high elevation species (right). Each box plot represents the median, interquartile ranges, maximum and minimum values of either CTmin or CTmax for each species. CTmin is shown by blue box plots and CTmax by orange box plots.

Another ANOVA was used to compare temperature ranges among populations. The analysis showed significant differences (One-way ANOVA,  $[F 7, 72] = 15.993$ ,  $p < 0.001$ ). *L. melanogaster* displayed the narrowest range of thermal tolerance, which differed from all other populations (Scheffe,  $p < 0.05$ ) but not from *L. sulphurophila* (Scheffe,  $p = 1.000$ ), *L. zonata* (Scheffe,  $p = 0.994$ ) and *L. versicolor* (Scheffe,  $p = 0.117$ ). Our data suggested that the most tolerant species to extreme temperatures were *L. perugiae*, *L. yaguajali*, and *L. vittata* (species distributed in low elevations) followed by *L. versicolor* and *L. dominicensis* (species which distribution extends into much higher elevations) since these two groups of species displayed broader ranges of temperature tolerance (Fig. 3) and their ranges did not differ significantly from each other (Scheffe,  $p > 0.05$ ).

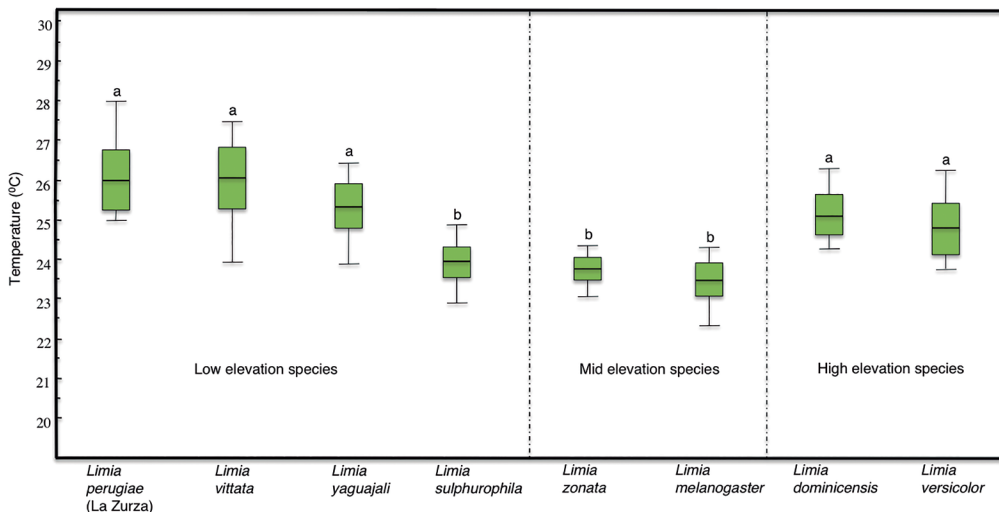


Figure 3. Box plots of the thermal ranges of the eight species of *Limia* included in this study. Species are grouped according to their elevation distribution in three groups: low elevation species (left), mid elevation species (center) and high elevation species (right). Each box plot represents the median, interquartile ranges, maximum and minimum values of either CTmin or CTmax for each species.

*Intra-specific analysis.* Population analyses for *L. perugiae* showed significant differences in CTmax (One-way ANOVA,  $F [3, 36] = 6.118, p = 0.02$ ) and CTmin (One-way ANOVA,  $F [3, 36] = 20.982, p < 0.001$ ). The population from Lake Enriquillo differed from the other three in both CTmax (Scheffe,  $p < 0.05$ ) and CTmin (Scheffe,  $p < 0.001$ ), (Fig. 4). There were also significant differences in ranges of thermal tolerance among populations of *L. perugiae* (One-way ANOVA,  $F [3, 36] = 3.409, p = 0.028$ ). In this case the population from Lake Enriquillo had significant differences with the population from Azufrada (Scheffe,  $p = 0.037$ ). This is surprising as the Azufrada population is also from Lake Enriquillo, just the north shore, not the south shore.

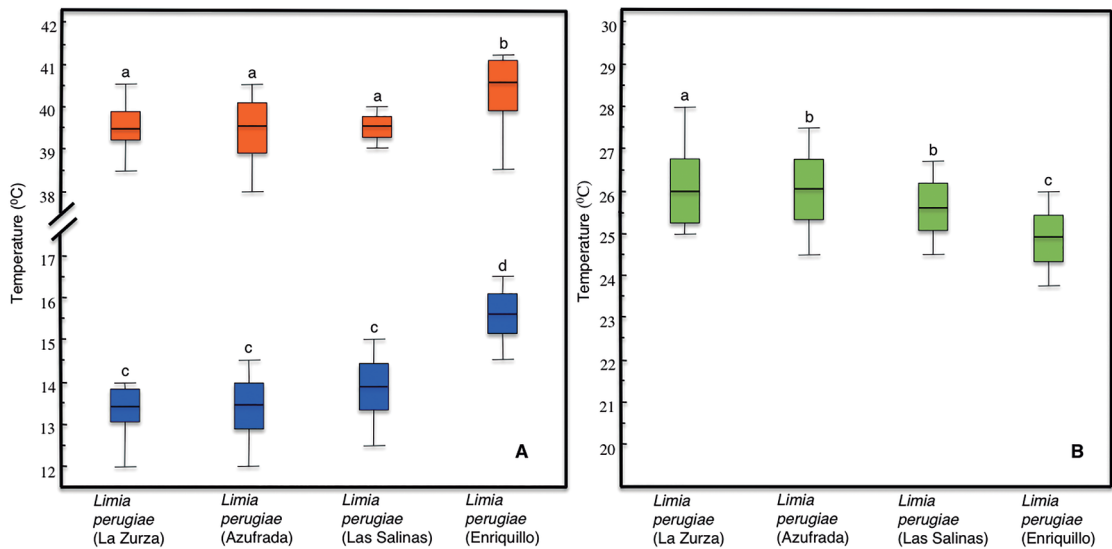


Figure 4. Thermal tolerances of four populations of *L. perugiae*. CTmin in blue box plots and CTmax in orange box plot (A), and box plots of thermal ranges of the four populations analyzed (B). Each box plot represents the median, interquartile ranges, maximum and minimum values.

## DISCUSSION

Our evidence suggests that thermal tolerance and altitudinal distribution of the populations of *Limia* species analyzed in this study are not related to temperature gradients expected according to Janzen hypothesis. The species studied here showed thermal tolerances not predicted by Janzen's hypothesis. Generally, even species from high altitudes (for the tropics) have broad thermal tolerances similar to species distributed in low elevations. Hence, the observed pattern does not separate the species as predicted.

Additionally, phylogeny did not explain species relationships according to thermal tolerance. However, failure to detect statistical significance may be due to the rather low number of species included in the analyses. Small sample sizes have been shown to influence the uncertainty and the expected values of most indices of phylogenetic signal, including the Pagel's lambda ( $\lambda$ ) (Munkemüller *et al.*, 2012). Powerful phylogenetic comparative analyses typically demand trait and phylogenetic data for over 50 species (Molina-Venegas & Rodriguez, 2017). The genus *Limia* only has 22 described species (Rodriguez-Silva *et al.*, 2020), which simply cannot satisfy the sampling requirements for robust macroevolutionary inferences.

Temperature tolerance ranges have been shown to shape species distributions and community compositions for some ectotherms in both tropical and temperate climates. Snyder and Weathers (1975) offered experimental evidence on the close relationship between the range of temperature tolerance and the environmental temperature variation in the distribution of several species of amphibians, showing that an increase in the environmental temperature variation also increases the range of temperature tolerance and consequently the distribution range of species. Estimating temperature tolerances and temperature ranges through the analysis of lower (CTmin) and upper (CTmax) thermal limits has been shown to be an efficient and useful method to assess species' capacity to acclimate to temperature changes in several ectotherms including terrestrial species (i.e. arthropods, reptiles and amphibians) and aquatic organisms (i.e. arthropods, mollusks and fish), (Van Berkum, 1988; Sunday *et al.*, 2011; Buckley & Huey, 2016). However, when using this methodology, the results can be influenced by experimental protocols and conditions in which either CTmin or CTmax are measured. Factors such as acclimation temperature of individuals being tested, the cooling or heating rate used, and the non-lethal endpoint chosen by the experimenter to determine CTmin or CTmax can influence the results (Lutterschmidt & Hutchison, 1997). Fortunately, though, the method has been in use for a long time, which has allowed testing and standardizing protocols for different animal groups. Hence, the technique offers repeatable and rapid quantitative measure of the thermal limits as well as predicts optimal temperate ranges of multiple species (Fischer & Schlupp, 2009; Kingsolver & Umbanhowar, 2018). Although specimens used in this experiment were kept in common garden conditions for different lengths of time, which might influence their tolerance to critical thermal limits; we standardized the acclimation time to a specific temperature range (25 °C – 27 °C) for 45 days under laboratory conditions. This acclimation time is considerably longer than others previously reported in studies of thermal limits in ectothermic organisms (Chanthy *et al.*, 2012; Moyano *et al.*, 2017; Tongnunui & Beamish, 2017), which ensures our data truly reflects the actual thermal tolerances of the species.

Janzen (1967) predicted that species occurring in high altitude in the tropics are specialized for lower temperatures than are low altitude species, which should be better adapted to cope with warmer temperatures. However, our results do not offer support for this prediction suggesting that factors other than temperature shape the distribution in populations of livebearing fishes of the genus *Limia*. In general, species and populations widespread in lowland habitats (*L. perugiae* populations from Hispaniola and *L. vittata* from Cuba) seem to be very tolerant to extreme temperatures (CTmax and CTmin) suggesting that they could also live in mountainous habitats. Given this, what could explain that none of these two species are found in high elevation streams either in Cuba or Hispaniola? We suggest that this is probably due to biotic factors such as competition. In the case of *L. vittata* in Cuba other dominant livebearing fishes (genera *Girardinus* and *Gambusia*) exploit available niches in mountain streams and on Hispaniola other *Limia* species and also species of *Poecilia* (*P. dominicensis*, *P. hispaniolana* and *P. elegans*) seem to be restricting *L. perugiae* to lowland environments. Evolutionary trade-offs between broad tolerance and competitive habitats have been shown to be common in different taxa. For instance, Robinson and Terborgh (1995) showed that interspecific aggression more than habitat suitability might explain spatial segregation patterns observed in Amazonian birds, and Griffis and Jaeger (1998) defined interspecific competition as cause of extinction of a species of salamander (*Plethodon shenandoah*) in the mountains of Shenandoah National Park, Virginia, USA.

Conversely, among species with distribution ranges that mostly include mid-elevations, there is a less consistent pattern in tolerance to extreme temperatures but with a general trend towards lower ranges of tolerance. Such are the cases of *L. melanogaster* from Jamaica and *L. zonata* from Hispaniola, which showed low tolerance ranges and were particularly

sensitive to low temperatures. Our recent exploratory work in the Caribbean has recorded these two species associated with permanent freshwater springs and spring runs that buffer temperature fluctuations. The two species abovementioned seem to be physiologically adapted to relatively narrow fluctuations in temperature, which may explain their limited tolerance range.

Another result of our study that runs counter to Janzen's hypothesis is that two strictly mountainous and locally distributed populations of the species *L. versicolor* and *L. dominicensis*, exhibited relatively broad tolerance ranges similar to species that typically are widespread distributed in low elevation environments. Increasing altitude is always accompanied by a decrease in annual average temperature (Sarmiento, 1986), which may indicate high elevation organisms are better adapted to cope with low temperatures. This general pattern was also present in our analysis where the highest study site, in this case Puerto Escondido with one population of *Limia dominicensis*, exhibited the lowest climate overlap index with the rest of the other sites. However, in tropical ecosystems species living at high elevations might benefit from evolving broad thermal tolerance to deal with diurnal changes in temperature (Ghalambor *et al.*, 2006), which in turn might explain why the two species also show a moderate tolerance for high temperatures. Temperature tolerances of high and mid elevation populations of *Limia* species relate to results of other studies in tropical amphibians (Navas, 1996; Navas *et al.*, 2013), which showed that species occurring in intermediate elevations were likely stenothermic given the relative thermal stability of those habitats. Conversely, species from higher elevations seemed to have evolved to lead with more changeable temperature (differences between day and night temperatures) and consequently develop broader tolerance ranges.

Our results also provide insights into local physiological adaptations of thermal tolerance within species, which suggests that conspecific populations in diverse habitats have somewhat independent evolutionary pathways (Snyder & Weathers, 1975). First, *L. perugiae* from the south shore of Lake Enriquillo, near where *L. sulphurophila* is found, differed from other three populations by showing a narrower thermal breadth. This result together with the lack of phylogenetic signal in thermal tolerance for species emphasizes the importance of biogeographical processes more than just phylogenetic patterns in analyses of climatic niche (Coelho *et al.*, 2019). Second, *L. sulphurophila*, a locally distributed species mainly known from sulfur springs on the southeastern shore of Lake Enriquillo in the Dominican Republic (Rivas, 1980), seems to be locally adapted to live in a high temperature sulfidic environment and is also physiologically adapted to resist high temperatures. This may prevent competitive exclusion as shown in fish species from temperate climates (Ohlberger *et al.*, 2008).

Our study has implications for conservation and is also pertinent in the context of climate change and species resilience to short-term temperature spikes. Our data provide evidence of species and populations that would be more vulnerable to temperature variation. In this case, the ones occurring in cool permanent freshwater springs in mid elevations and *L. sulphurophila* (a local endemic species) seemed to be more susceptible to temperature fluctuations because of their narrower thermal breadth. Another issue that may affect conservation of native *Limia* species is the introduction of invasive livebearing fishes, such as *Poecilia reticulata* (Guppy). This species has recently been reported as one of the most tolerant ornamental fish to extreme temperatures (Yanar *et al.*, 2019), which may be additional evidence of the invasive success of guppies and in some extent explain why this species becoming dominant in tropical ecosystems.

The implications of temperature for fish physiology and fitness (Niehaus *et al.*, 2012; Payne *et al.*, 2016) make the analysis of thermal limits particularly important in determining distribution of fishes (Culumber *et al.*, 2012). Even though our study does not provide a

comprehensive test for Janzen's hypothesis, it presents evidence at local scales to analyze how elevational gradients may affect the distribution of freshwater fishes, which is a barely studied zoological group in the Caribbean. While it does not offer evidence supporting Janzen's predictions about climatic variation across elevations, physiological adaptation and species distribution for this group of fish; the study emphasizes the importance of testing the validity of Janzen's mountain passes hypothesis across multiple taxa. In addition, like previous studies this work stresses the significance of other factors such as species interactions, diet specializations, and even thermoregulatory behavior [as shown by Muñoz & Bodensteiner (2019) in Caribbean anoles] when interpreting current altitudinal distribution patterns of species.

#### ACKNOWLEDGEMENTS

This study was supported by the National Geographic Society (WW-054R-17) and the University of Oklahoma. We would like to thank the governments and corresponding ministries of Jamaica and the Dominican Republic for kindly issuing collecting permits. In addition, we thank to the Museo Nacional de Historia Natural "Prof. Eugenio de Jesus Marcano" in Santo Domingo, Dominican Republic and especially to Patricia Torres Pineda and Carlos Suriel. We also thank Carlos Rodriguez from the Ministerio de Educación Superior, Ciencia y Tecnología in the Dominican Republic for his support. Thanks to Ricardo Betancur and Emanuell Ribero their insights and for helping with the analysis of phylogenetic signal. Thanks to Caryn Vaughn, Laura Stein and Bruce Hoagland for their advice and comments on the manuscript. We are grateful to Carolyn Burt, Kerri-Ann Bennett, Stephan Bräger, Heriberto Encarnación Lara, Kenia Ng Alvarado, and Marcos José Rodriguez for help with fieldwork. Trai Spikes, Sophie Huebler, Margaret Zwick, Zeeshawn Beg, and Nabiha Ahmad helped with fish care. Gabriel Costa helped with statistical analysis. Finally, we are grateful to the reviewers for constructive comments on the manuscript.

#### LITERATURE CITED

- Arnaudo, M. E., N. Toledo, L. Soibelzonand, & P. Bona. 2019. Phylogenetic signal analysis in the basicranium of Ursidae (Carnivora, Mammalia). *PeerJ*:e6597. <https://doi.org/10.7717/peerj.6597>
- Beitinger, T. L., W. A. Bennett, & R. W. McCauley. 2000. Temperature tolerances of North American freshwater fishes exposed to dynamic changes in temperature. *Environmental Biology of Fishes*, 58: 237–275.
- Buckley, L. B., & R. B. Huey. 2016. Temperature extremes: geographic patterns, recent changes, and implications for organismal vulnerabilities. *Global Change Biology*, 22: 3829–3842. <https://doi:10.1111/gcb.13313>
- Burgess, G. H., & R. Franz. 1989. Zoogeography of the Antillean freshwater fish fauna. In: Woods, C. A. & F. E. Sergile (Eds) *Biogeography of the West Indies: Patterns and Perspectives*. CRF Press, Boca Raton FL, 263–304.
- Carvajal-Quintero, J. D., F. Escobar, F. Alvarado, F. A. Villa-Navarro, U. Jaramillo-Villa, & J. A. Maldonado-Ocampo. 2015. Variation in freshwater fish assemblages along a regional elevation gradient in the northern Andes, Colombia. *Ecology and Evolution*, 5 (13): 2608–2620.

- Chanthy, P., R. J. Martin, R. V. Gunning, & N. R. Andrew. 2012. The effects of thermal acclimation on lethal temperatures and critical thermal limits in the green vegetable bug, *Nezara viridula* (L.) (Hemiptera: Pentatomidae). *Frontiers in Physiology*, 3: 465. <https://doi.org/10.3389/fphys.2012.00465>
- Coelho, M. T. P., J. F. B. Rodrigues, J. A. F. Diniz-Filho, & T. F. Rangel. 2019. Biogeographical history constrains climatic niche diversification without adaptive forces driving evolution. *Journal of Biogeography*, 46: 1020–1028. <https://doi.org/10.1111/jbi.13553>
- Cowles, R. B., & C. M. Bogert. 1944. A preliminary study of the thermal requirements of desert reptiles. *Bulletin of the American Museum of Natural History*, 83: 265–296.
- Culumber, Z. W., D. B. Shepard, S. W. Coleman, G. G. Rosenthal, & M. Tobler. 2012. Physiological adaptation along environmental gradients and replicated hybrid zone structure in swordtails (Teleostei: *Xiphophorus*). *Journal of Evolutionary Biology*, 25: 1800–1814.
- Fischer, C., & I. Schlupp. 2009. Differences in thermal tolerance in coexisting sexual and asexual mollies (*Poecilia*, Poeciliidae, Teleostei). *Journal of Fish Biology*, 74: 1662–1668. <https://doi.org/10.1111/j.1095-8649.2009.02214.x>
- Ghalambor, C. K., R. B. Huey, P. R. Martin, J. J. Tewksbury, & G. Wang. 2006. Are mountain passes higher in the tropics? Janzen's hypothesis revisited. *Integrative and Comparative Biology*, 16 (1): 5–17. <https://doi.org/10.1093/icb/icj003>
- Gilbert, P. S., J. Wu, M. W. Simon, J. S. Sinsheimer, & M. E. Alfaro. 2018. Filtering nucleotide sites by phylogenetic signal to noise ratio increases confidence in the Neoaves phylogeny generated from ultraconserved elements. *Molecular Phylogenetics and Evolution*, 126: 116–128. <https://doi.org/10.1016/j.ympev.2018.03.033>
- Gingras, B., E. Mohandesan, D. Boko, & W. F. Tecumseh. 2013. Phylogenetic signal in the acoustic parameters of the advertisement calls of four clades of anurans. *BMC Evolutionary Biology*, 13: 134. <https://doi.org/10.1186/1471-2148-13-134>
- Graham, C. H., A. C. Carnaval, C. D. Cadena, K. R. Zamudio, T. E. Roberts, & J. L. Parra. 2014. The origin and maintenance of montane diversity: integrating evolutionary and ecological processes. *Ecography*, 37: 711–719. <https://doi.org/10.1111/ecog.00578>
- Griffis, M. R., & R. G. Jaeger. 1998. Competition leads to an extinction-prone species of salamander: interspecific territoriality in a metapopulation. *Ecology*, 79: 2494–2502.
- Hamilton, A. 2001. Phylogeny of *Limia* (Teleostei: Poeciliidae) based on NADH dehydrogenase subunit 2 sequences. *Molecular Phylogenetics and Evolution*, 19 (2): 277–289. <https://doi.org/10.1006/mpev.2000.0919>
- Hijmans, R. J. 2020. Raster: Geographic Data Analysis and Modeling. R package version 3.0–12. <https://CRAN.R-project.org/package=raster> (accessed: 06/20/2020).
- Hua, X. 2016. The impact of seasonality on niche breadth, distribution range and species richness: a theoretical exploration of Janzen's hypothesis. *Proceedings Biological Sciences*, 283 (1835): 20160349. <https://doi.org/10.1098/rspb.2016.0349>

- Janzen, D. H. 1967. Why mountain passes are higher in the tropics. *American Naturalist*, 101: 233–247. <https://doi.org/10.1086/282487>
- Jaramillo-Villa, U., J. A. Maldonado-Ocampo, & F. Escobar. 2010. Altitudinal variation in fish assemblage diversity in streams of the central Andes of Colombia. *Journal of Fish Biology*, 76: 2401–2417. <https://doi.org/10.1111/j.1095-8649.2010.02629.x>
- Kamilar, J. M., & N. Cooper. 2013. Phylogenetic signal in primate behaviour, ecology and life history. *Philosophical Transactions of the Royal Society of London Biological Sciences*, 368: 20120341. <http://dx.doi.org/10.1098/rstb.2012.0341>
- Kingsolver, J. G., & J. Umbanhowar. 2018. The analysis and interpretation of critical temperatures. *Journal of Experimental Biology*, 2018 (221). <https://jeb167858>. doi:10.1242/jeb.167858
- Layne, J. R. Jr., & D. L. Claussen. 1982. The time courses of CTMax and CTMin acclimation in the salamander *Desmognathus fuscus*. *Journal of Thermal Biology*, 7 (3): 139–141.
- Leiva, F. P., P. Calosi, & W. C. E. P. Verberk. 2019. Scaling of thermal tolerance with body mass and genome size in ectotherms: a comparison between water– and air–breathers. *Philosophical Transactions of the Royal Society of London Biological Sciences*, 374: 20190035. <http://dx.doi.org/10.1098/rstb.2019.0035>
- Lowe, C. H., & V. J. Vance. 1955. Acclimation of the critical thermal maximum of the reptile *Urosaurus ornatus*. *Science*, 122: 73–74.
- Lutterschmidt, W. I., & V. H. Hutchison. 1997. The critical thermal maximum: data to support the onset of muscle spasm as the definitive end point. *Canadian Journal of Zoology*, 75: 1553–1560.
- McCain, C. M. 2009. Vertebrate range sizes indicate that mountains may be ‘higher’ in the tropics. *Ecology Letters*, 12: 550–560. <https://doi.org/10.1111/j.1461-0248.2009.01308.x>
- Molina-Venegas, R., & M. A. Rodriguez. 2017. Revisiting phylogenetic signal; strong or negligible impacts of polytomies and branch length information? *BMC Evolutionary Biology*, 17 (53): 1–10.
- Moyano, M., C. Candebat, Y. Ruhbaum, S. Alvarez-Fernandez, G. Claireaux, J. L. Zambonino-Infante, & M. A. Peck. 2017. Effects of warming rate, acclimation temperature and ontogeny on the critical thermal maximum of temperate marine fish larvae. *Plos One*, 12 (7): e0179928. <https://doi.org/10.1371/journal.pone.0179928>
- Munkemüller, T., S. Lavergne, B. Bzeznik, S. Dray, T. Jombart, K. Schiffers, & W. Thuiller. 2012. How to measure and test phylogenetic signal. *Methods in Ecology and Evolution*, 3: 743–756. <https://doi.org/10.1111/j.2041-210X.2012.00196.x>
- Muñoz, M. M., J. E. Wegener, & A. C. Algar. 2014. Untangling intra– and interspecific effects on body size clines reveals divergent processes structuring convergent patterns in *Anolis* lizards. *American Naturalist*, 184: 636–646.
- Muñoz, M. M., & B. L. Bodensteiner. 2019. Janzen’s hypothesis meets the Bogert effect: Connecting climate variation, thermoregulatory behavior and rates of physiological evolution. *Integrative Organismal Biology*, 1–12. <https://doi.org/10.1093/iob/oby002>

- Navas, C. A. 1996. Implications of microhabitat selection and patterns of activity on the thermal ecology of high elevation neotropical anurans. *Oecologia*, 108: 617–626. <https://doi.org/10.1007/BF00329034>
- Navas, C. A., J. M. Carvajalino-Fernández, L. P. Saboyá-Acosta, L. A. Rueda-Solano, & M. A. Carvajalino-Fernández. 2013. The body temperature of active amphibians along a tropical elevation gradient: Patterns of mean and variance and inference from environmental data. *Functional Ecology*, 27: 1145–1154. <https://doi.org/10.1111/1365-2435.12106>
- Niehaus, A. C., M. J. Jr. Angilletta, M. W. Sears, C. E. Franklin, & R. S. Wilson. 2012. Predicting the physiological performance of ectotherms in fluctuating thermal environments. *Journal of Experimental Biology*, 215: 694–701. <https://doi.org/10.1242/jeb.058032>
- Ohlberger, J., T. Mehner, G. Staaks, & F. Hölker. 2008. Temperature-related physiological adaptations promote ecological divergence in a sympatric species pair of temperate freshwater fish, *Coregonus* spp. *Functional Ecology*, 22: 501–508.
- Pagel, M. 1994. Detecting correlated evolution on phylogenies: a general method for the comparative analysis of discrete characters. *Proceedings of the Royal Society of London Biological Sciences*, 255: 37–45. <https://doi.org/10.1098/rspb.1994.0006>
- Payne, N. L., J. A. Smith, D. E. van der Meulen, M. D. Taylor, Y. Y. Watanabe, A. Takahashi, T. A. Marzullo, C. A. Gray, G. Cadiou, & I. M. Suthers. 2016. Temperature dependence of fish performance in the wild: links with species biogeography and physiological thermal tolerance. *Functional Ecology*, 30 (6): 903–912. <https://doi.org/10.1111/1365-2435.12618>
- Pintanel, P., M. Tejedo, S. R. Ron, G. A. Llorente, & A. Merino-Viteri. 2019. Elevational and microclimatic drivers of thermal tolerance in Andean *Pristimantis* frogs. *Journal of Biogeography*, 46: 1664–1675. <https://doi.org/10.1111/jbi.13596>
- Polato, N. R., B. A. Gill, A. A. Shah, M. M. Gray, K. L. Casner, A. Barthelet, & K. R. Zamudio. 2018. Narrow thermal tolerance and low dispersal drive higher speciation in tropical mountains. *Proceedings of the National Academy of Sciences*, 115: 12471–12476. <https://doi.org/10.1073/pnas.1809326115>
- R Core Team. 2013. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.
- Revell, L. J. 2012. Phytools: an R package for phylogenetic comparative biology (and other things). *Methods in Ecology and Evolution*, 3: 217–222. <https://doi.org/10.1111/j.2041-210X.2011.00169.x>
- Rivas, L. R. 1980. Eight new species of poeciliid fishes of the genus *Limia* from Hispaniola. *Northeast Gulf Science*, 4 (1): 28–38.
- Robinson, S. K., & J. Terborgh. 1995. Interspecific aggression and habitat selection by Amazonian birds. *Journal of Animal Ecology*, 64: 1–11.
- Rodríguez, C. M. 1997. Phylogenetic analysis of the tribe Poeciliini (Cyprinodontiformes: Poeciliidae). *Copeia*, 1997 (4): 663–679.



- Rodriguez-Silva, R., P. Torres-Pineda, & J. Josaphat. 2020. *Limia mandibularis*, a new livebearing fish (Cyprinodontiformes: Poeciliidae) from Lake Miragoane, Haiti. *Zootaxa*, 4768 (3): 395–404. <https://doi.org/10.11646/zootaxa.4768.3.6>
- Rodriguez, R. S., P. Torres-Pineda, C. M. Rodriguez, & I. Schlupp. 2020. Distribution range extension of Yaguajal Limia, *Limia yaguajali* (Teleostei: Poeciliidae) from north of the Dominican Republic, Hispaniola. *Novitates Caribaea*, 15: 127–133.
- Sarmiento, G. 1986. Ecological features of climate in high tropical mountains. (11–46). In: Vuilleumier, F., & M. Monasterio (Eds.). *High altitude tropical biogeography*. Oxford University Press, New York, 649 pp.
- Snyder, G. K., & W. W. Weathers. 1975. Temperature adaptations in amphibians. *American Naturalist*, 109: 93–101.
- Spotila, J. R. 1972. Role of temperature and water in the ecology of lungless salamanders. *Ecological Monographs*, 42 (1972): 95–125.
- Sunday, J. M., A. E. Bates, & N. K. Dulvy. 2011. Global analysis of thermal tolerance and latitude in ectotherms. *Proceedings of Biological Sciences*, 278: 1823–1830.
- Tongnunui, S., & F. W. H. Beamish. 2017. Critical thermal maximum, temperature acclimation and climate effects on Thai freshwater fishes. *Environment Asia*, 10 (1): 109–117.
- Valdivieso, D., & J. R. Tamsitt. 1974. Thermal relationships of the neo-tropical frog *Hyla labialis* (Anura: Hylidae). *Life Sciences Occasional Papers Royal Ontario Museum*, 26: 1–10.
- Van Berkum, F. H. 1988. Latitudinal patterns of the thermal sensitivity of sprint speed in lizards. *American Naturalist*, 132: 327–343.
- Weaver, P. F., O. Tello, J. Krieger, A. Marmolejo, K. F. Weaver, J. V. Garcia, & A. Cruz. 2016a. Hypersalinity drives physiological and morphological changes in *Limia perugiae* (Poeciliidae). *Biology Open*, 5: 1093–1101.
- Weaver, P. F., A. Cruz, S. Johnson, J. Dupin, & K. F. Weaver. 2016b. Colonizing the Caribbean: biogeography and evolution of livebearing fishes of the genus *Limia* (Poeciliidae). *Journal of Biogeography*, 43: 1808–1819. <https://doi.org/10.1111/jbi.12798>
- Wollenberg, K. C., I. J. Wang, R. E. Glor, & J. B. Losos. 2013. Determinism in the diversification of Hispaniolan trunk-ground anolis (*Anolis cybotes* species complex). *Evolution*, 67: 3175–3190. <https://doi.org/10.1111/evo.12184>
- Yanar, M., S. Erdoğan, & M. Kumlu. 2019. Thermal tolerance of thirteen popular ornamental fish species. *Aquaculture*, 501: 382–386. <https://doi.org/10.1016/j.aquaculture.2018.11.041>